DIVISION OF AGRICULTURE
RESEARCH & EXTENSION
University of Arkansas System

Sherrie Smith

Ricky Corder

SPDN
Southern Plant Diagnostic Network

Department of Plant Pathology

PLANT HEALTH

CLINIC NEWS

Issue 7, April 18, 2016

This bulletin from the Cooperative Extension Plant Health Clinic (Plant Disease Clinic) is an electronic update about diseases and other problems observed in our lab each month. Input from everybody interested in plants is welcome and appreciated.

Camellia

Tea scale, *Fiorinia theae,* is a major pest of camellias as well as holly, tea, citrus, dogwood, bottlebrush, kumquat, mango, and olive among others. These insects have piercing mouthparts that enable them to feed on the sugary contents of plant cells. This causes a yellow stippling of the upper leaf surface. Heavy infestations may cause premature leaf dropping, decline in the health of the plant, and occasionally even death. Tea scale primarily infests the underside of the leaves, making spraying for them more difficult. Adding to the difficulty is that Tea scale has multiple generations in the mid-south. Sprays of fine horticultural oils are effective against Tea scale if good coverage is achieved. Systemic insecticides such as Bayer Advanced Insect Control for trees and shrubs may also be used.

Camellia Tea scale-*Fiorinia theae*

Russell Parker University of Arkansas Cooperative Extension

Camellia Tea scale-*Fiorinia theae*

Russell Parker University of Arkansas Cooperative Extension

Peach/nectarine

Spores from the fungus *Taphrina deformans* overwinters on twigs and bud scales. Infection occurs at bud break early in the spring during cool, wet weather. Blister-like swellings, curling, thickening, puckering, and discoloration of the leaves are the first symptoms of Peach leaf curl. Affected areas may turn pink, red or yellow. In severe cases, defoliation occurs along with substantial yield loss. Peach leaf curl is easily controlled with one well-timed fungicide application in the fall after 90% of the leaves have dropped, or very early in the

DIVISION OF AGRICULTURE
RESEARCH & EXTENSION
University of Arkansas System

Sherrie Smith

Ricky Corder

SPDN
Southern Plant Diagnostic Network

Department of Plant Pathology

PLANT HEALTH

CLINIC NEWS

Issue 7, April 18, 2016

spring before the buds begin to swell. Chlorothalonil or copper sprays are effective. It is too late for chemical control this spring, but if only a few leaves are infected, they may be handpicked and destroyed to reduce inoculum levels.

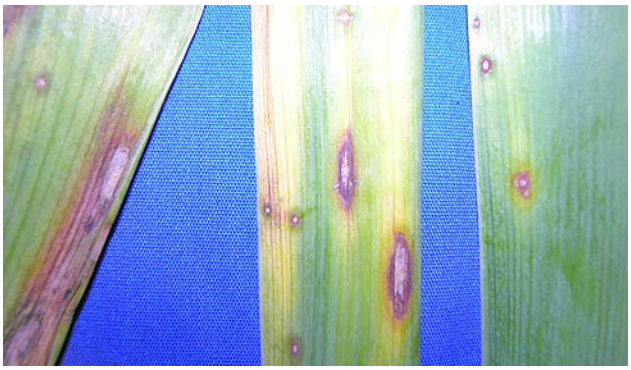
Peach leaf curl-*Taphrina deformans*

Michael Sullivan, former University of Arkansas Cooperative Extension Agent

Iris

The bearded Irises are blooming and filling the air with their delightful fragrance. Irises are dependable, hardy plants that get by with a minimum of care. They are less prone to disease when planted in full sun in well-drained, rich loam soil. A pH of 6.0-7.0 is preferred. However, during periods of prolonged warm, wet weather, they may be susceptible to Iris Leaf spot, caused by *Mycosphaerella macrospora,* formerly *Didymellina macrospora*. Symptoms begin on the leaves as tiny, green to yellow, water-soaked spots, which become oval brown lesions with water-soaked yellow margins. After bloom, the spots enlarge to form large, irregular, dead areas. Old lesions become gray with reddish brown to dark brown borders. Tip dieback and leaf curl are common. Severely affected leaves may die completely. If this happens frequently, it weakens the plant and reduces bloom quality. Daylily, freesia, gladiolus, and narcissus are also susceptible. Good cultural practices can help greatly in reducing this disease. All iris debris should be cleaned up in the fall, or before new leaves appear in the spring. During the growing season, diseased portions of the leaf should be removed from the plant and disposed of away from the planting. Overly crowded clumps should be divided and replanted in the fall. Sprinkler irrigation should be avoided and plants watered at ground level. Fungicide sprays may be applied when the new fan leaves are four to six inches, and repeated four or five times at 7 to 10 day intervals. Products containing chlorothalonil, or myclobutanil, or thiophanate-methyl, or mancozeb, or trifloxystrobin are effective. A spreader sticker should be added to enable the fungicide to stick to the waxy iris leaves.

Iris leaf spot- *Mycosphaerella macrospora (*syn. *Didymellina macrospora)*



Sherrie Smith University of Arkansas Cooperative Extension

DIVISION OF AGRICULTURE
RESEARCH & EXTENSION
University of Arkansas System

Sherrie Smith

Ricky Corder

SPDN
Southern Plant Diagnostic Network

Department of Plant Pathology

PLANT HEALTH

CLINIC NEWS

Issue 7, April 18, 2016

Rhododendron

A common leaf spot disease of rhododendron is Cercospora leaf spot, caused by the fungus *Cercospora handelii.* Dark brown, angular leaf spots occur on the upper leaf surface. Lesions on hybrid cultivars may have silvery centers as a result of the separation of the leaf epidermis. The most damage occurs in crowded plantings with poor air circulation. Lower foliage is usually affected the worse. Good sanitation, proper spacing, and avoidance of overhead irrigation are useful for control of Cercospora leaf spot. Homeowners may use Spectracide Immunox, or Fertilome Liquid Systemic Fungicide, or Ortho Max Garden Disease Control, or Fertilome Liquid Fungicide, or Garden Tech Daconil Fungicide Conc., or Green Light Fung-Away Fungicide, or Bonide Fung-onil Multipurpose Fungicide, or Green Light Systemic Fungicide, or Fertilome Halt Systemic, or Ortho Rose Pride Rose & Shrub Disease Control, or Bayer Advanced Garden-Disease Control for Roses, Flowers, Shrubs.

Rhododendron leaf spot-
Cercospora handelii

Sherrie Smith University of Arkansas Cooperative Extension

Rhododendron leaf spot-
Cercospora handelii

Sherrie Smith University of Arkansas Cooperative Extension

DIVISION OF AGRICULTURE
RESEARCH & EXTENSION
University of Arkansas System

Sherrie Smith

Ricky Corder

SPDN
Southern Plant Diagnostic Network

Department of Plant Pathology

PLANT HEALTH

CLINIC NEWS

Issue 7, April 18, 2016

Request for help from Dr. Robbins:

Root knot nematode populations are needed for our Arkansas species study. I am a nematologist in the department of Plant Pathology in Fayetteville. My student and I are trying to amass populations of as many species of Root knot nematode (Meloidogyne sp.) as possible for species identification using molecular techniques. At present no root knot species in Arkansas have been identified using molecular technology. We are interested in receiving populations from home gardens, shrubs, flowers, trees and grasses. For samples we need about a pint of soil and feeder roots in a sealed plastic bag that is plainly identified by plant host, location (City County, physical address, collector and date of collection). Please send samples to us at the follow address:

Dr. Robert Robbins
Cralley-Warren Research Center
2601 N. Young Ave
Fayetteville, AR 72701
Phone 479-575-2555
Fax 479-575-3348
Email: rrobbin@uark.edu